FISH KILL INVESTIGATION REPORT FORM 1 Date: 2 Time of Arrival: 3. Waterbody Location: 4. **Person reporting:** Name: Phone: Total Time spent at site: Address: Affiliation: 5. # of fish Killed: 6. Dimensions of fish kill: 7. Fish Species Affected: Fish Size Incident Size: Same Different Range ____ to ___ in. Minor <100 Same Different Range ____ to ___ in. _____ by ____ Range ____ to ___ in. Same Different Range ____ to ___ Maior >1000 Same Different in 9. Weather Range ____ to ___ in. 8. Fish Species Not Affected Same Different Temp (F) Same □ Different Range to in. 7a. Other Species Affected: Cloud Cover (%) Dying \square Lethargic Dead □ I ive □ Precipitation (%) 2. Dead □ Dying \Box Live □ Wind Speed (mph) Lethargic Dead □ Dying \Box Live □ Wind direction Dead Dving Lethargic Live 12. Fish Condition: 10. Water Quality: 11. Water Condition: Discoloration Temp (C): _____ □ Increased respiration □ Emaciated Dying Gills flared Red/pink gills pH: _____ DO: ____ Odd fin position Turbid □ Eyes sunken in □ Spasms, convulsions Sediment Loading Colored: _____ Conductivity: Gill clubbing Salinity: Odor: _____ Tidal Stage: _____ Excessive mucus Trying to get Chlorine: _____ Lesions П П SAV/ macroalgae Run samples for: Other П 13. Symptoms/Conditions **Possible Cause** Possible Source Source present? Sewage Treatment Plan Yes □ No □ Fish coming to surface gulping for air Livestock Feedlot Yes □ No □ Low dissolved oxygen Oxygen depletion Irrigation/De-icing Runoff Yes □ No 🗆 Decaying Plant Matter Yes ⊓ No \Box Dying Algal Bloom Yes □ No □ Ammonia Chemicals Early oxygen depletion Yes □ No 🗆 Fish coming to surface gulping for air with slow re-oxygenation Adequate dissolved oxygen Livestock Feedlot Yes □ No □ Heavy Metal Plant Yes □ No 🗆 Fish swimming erratically Chemical pollution Chemical Waste Facility Yes □ No 🗆 Sewage Treatment Plant Yes □ No □ Farms. Crop fields Yes □ No 🗆 Pesticide. herbicide Aerial Crop Sprayer Yes 🗆 No \square Fish dying or dead after heavy rain washed out/runoff Man/mechanical Sprayer No 🗆

Oxygen depletion

Turbines or thermal shock

Spawning stress, disease

Sent to: _____

Acid

Additional	Comments:

Photos taken

14. Documentation and Samples:

Fish coming to surface gulping for air

• Fish dying below a dam or industrial plant

• Kill restricted to one species or size class

• Low pH □ Good clarity □ Orange Discoloration

Water samples

Number: _____

Fish Samples

Number: _____

Dredging/ Marina activity

Coal/Strip Mining

Pathogens, WQ poor

Heated water

Yes □

Yes □

Yes 🗆

Yes □

Yes 🗆

Tested For: Tested For:

No □

No □

No □

No 🗆

15. Prepared By:

Fish Kill/Incident Notification

Date of Kill/Incident:					
Date Reported:	Reported: Time Reported:				
Name of Reporter:					
Address:		Phone:			
Organization Associated	With:				
Water(s) Involved:					
Specific Location (bridge, highw					
Suspected Reason For Fish Kill/	incident (natural / po	ollution):			
Location of Source:					
Name of Alleged Polluter (if app	licable):				
Address:		Phone:			
Species Involved:					
Fish Affected? Yes _	No				
Approximate Number:	Still Dying	? Yes	No Some → ~%		
Additional Comments:					
Persons and Agencies Notifie	d To Respond:				
<u>NAME</u>	DATE/TIME	PHONE	REPORT SENT TO		
1	_		Yes No		
2		_	Yes No		
Division of Enforcement Notified	at (401) 222-3070	Yes	No		
Report Prepared By:		Further Action N	Needed?Yes No		

FISH KILL RESPONSE EQUIPMENT CHECKLIST

<u>In kit</u>	Need to locate	
		1. Fish kill forms
		2. Waterproof field notebook
		3. ID tags (COC tags)
		4. Clipboard
		5. Pens
		6. Permanent markers
		7. Sampling bottles
		8. Dissecting/Necropsy kit
		9. First aid kit
		10. Tape measure
		11. Disposable chemical resistant gloves
		12. Fish shipping bags & ties
		13. Garbage bags
		14. Paper towels
		15. Measuring board
		16. Hand tally's/counters
		17. Camera
		18. Cooler
		19. lce
		20. Air pumps
		21. Oxygen tablets
		22. YSI meter (DO,salinity,temp,conductivity)
		23. LaMotte Saltwater Test Kit
		24. Chlorine test strips
		25. Thermometer
		26. Waders/boots
		27. AFS Guidelines/Fish Inv. Manual
		28. Fish tote
		29. Dip nets
		30. Scale
		31. Flashlight
		32. Nautical chart

FISH-COUNTING RECORD

		<u></u>	ISH-COUNTING	RECORD		
Date:	Time:	Start	Finish	Name of investi	gator(s):	
	aterbody Name:					
	# of Transects					
			SPECIES			
CHES						
01120						
1						
2						
3						
4						
5						
6						
7						
8						
9						
10						
11						
12						
13						
14						
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R.I. Department of Environmental Management

Division of Fish & Wildlife Fort Wetherill Marine Laboratory 3 Fort Wetherill Road Jamestown, Rhode Island 02835

Fish Kill Response - Standard Operating Procedure (SOP)

This SOP is a guide for Marine Fisheries staff in responding to reports of fish kills, and will be applicable to all fish, shellfish and other invertebrates found within Rhode Islands marine/estuarine waters. (See also the <u>Protocol For DEM Response To a Report of an Environmental Incident On Narragansett Bayy</u>.)

Definition of a fish kill:

A 'fish kill' is a significant and sudden death of fish, shellfish and other aquatic animals. Such events are characterized by large numbers of animals dying over a short time, usually in a defined area.

Initial Notification

- 1. When a call is received, the staff member who receives the call should fill out the *Fish Kill Notification Form*.
- 2. If a call is received from a private party, contact DEM Emergency dispatch at 222-3070 so information can be routed to the proper office(s).
- 3. Based on the initial report, proper staff should be notified to respond.
- 4. The number of staff to respond will be determined by the size/severity of the kill.
- 5. Responding staff should locate proper forms, equipment and ice (located in dry lab freezer), and proceed to the site. (Refer to the <u>Fish Kill Response Equipment Checklist</u> for a full list of equipment and the *Fish Kill Investigation Package* for a complete set of forms.)

Once on location:

- 1. Talk to any witnesses/observers. Take a statement from any person at the scene who may have pertinent information. Their identity should be established, and statements should be signed and witnessed.
- 2. Any information recorded during an inspection should be written in pen. Do NOT use pencil!
- 3. Determine the extent of the kill by walking the length of shoreline or accessing the site by boat.
- 4. If visual observations confirm an obvious chemical contaminant, immediately contact the DEM Office of Emergency Response.
 - During normal business hours (Mon-Fri, 8-4), call (401) 222-1360. Otherwise -- off-hours or if unsuccessful reaching that number -- call the DEM Division of Law Enforcement at (401) 222-3070.
 - If there is chemical contamination, do NOT sample the area! Wait for the DEM Office of Emergency Response to provide further instructions.

5. If no chemical contamination is observed, proceed with an investigation. Begin to fill out <u>The Fish Kill Investigation Report Form</u> and continue to use it throughout site inspection. Information should include, but is not limited to:

- # of fish - Symptoms/ conditions

WeatherSpecies affectedWater quality

- 6. Photographs should be taken of the entire site. Evidence of dead or affected fish and any other materials suspected of being associated with the fish kill should also be photographed. The date, time and location of sequential photographs, and the name of the photographer should be recorded in the field notebook.
- 7. Take water quality samples along the entire extent of the kill. Sampling should be conducted in the kill zone and in unimpacted areas. Document the exact location of each sample in the field notebook.
 - a. Water samples for analysis should be taken in clean plastic *Nalgene* bottles.
 - b. The bottle should be held underwater with its cap on until the desired depth is reached.
 - c. Proceed to remove the cap and fill the bottle up to its neck.
 - d. The sample bottle should now be sealed without retaining an air bubble.
 - e. Place bottles on ice in a cooler.
- 8. If possible, control or background conditions should be sampled using the same methods as described above. (Control samples capture water quality conditions outside of the kill zone.)
- 9. Instruments should be calibrated before sampling and should be documented in field notes. YSI readings (temp, DO, salinity, conductivity) should be taken immediately when arriving at the fish kill site for the most accurate representation of water quality. All data should be recorded in field notebook and should be dated and signed.
- 10. When ready to take fish samples or handle fish, wear proper protection (gloves). If fish are alive, make every attempt to keep a few of them alive in ambient water. Do NOT place fish in "fresh" saltwater.
- 11. If no fish are alive, then place 3-5 of the freshest dead fish in separate bags and cover with bags of ice. Make sure each bag is labeled with a chain of custody tag. Do not place fish directly on ice!! Keep them in a cooler. Further investigation will determine whether water and/or fish samples should be sent to a lab for further evaluation.
- 12. Make physical observations, and if possible perform a simplified necropsy on a few fish. Refer to the *Fish Kill Investigation Form* for a list of possible physical conditions. Look for any abnormalities, which could lead to a possible explanation of the fish kill. Record any relevant information in field notebook and data sheets. Photographs SHOULD be taken of dead or affected fish and any other affected organisms!!!!!
- 13. Based on the type of kill and/or location, the most accurate counting method should be determined by using one of the following:

a. Shoreline Count

- Determine the length of the affected shoreline by subdividing the shoreline into equal segments of a fixed length.
- Count the total number of segments in the affected shoreline.

- Then randomly select at least 3 segments.
- Multiply the average or total count of fish in each segment by the appropriate expansion factor.
- Hence: (# of fish/segment) x (total number of segments in fish kill zone) = Estimate of total population killed

b. Area sampling (lakes)

- Total Number = (# fish counted) x EV
- EV = (Total area effected)/(area sampled)
- For example: 100 fish were counted in an area of 10 acres². The lake was 100 acres². Hence: Total number = 100 x (100/10) = 1000 fish

c. Complete Count

- Count all fish over the entire extent of the kill. (This method is appropriate for a relatively small kill.)
- 14. Regardless of the counting method, staff should identify, measure and weigh (if appropriate), all fish within sampling area(s). Only fish larger than 6 inches should be weighed.
- 15. Record all information on the *Fish Counting Record Form*.
- 16. If deemed necessary by field staff, begin to measure appropriate water quality parameters by using the *Saltwater LaMotte Kit* and other test kits. Record all information in a field notebook and on correct forms.
- 17. When all water quality testing is complete, a field biologist should determine whether fish and/or water samples should be brought to a laboratory for further testing.
- 18. If the field biologist determines that further testing is needed, contact the appropriate laboratory for proper instructions and handling methods. Appropriate laboratories include:

Pathology

URI-East Farm, Pathology (fish/shellfish)
 Contact: Marta Gomez-Chiarri, Pathologist (401) 874-2917

Toxicology

1. RIDOH Laboratory, Toxicology & Microbiology Contact: Dr. Ewa King, (401) 222-1999 (toxicology) Contact: Dr. Ken Jones, (401) 222-5596 (microbiology)

2. ESS Laboratory, Toxicology Contact: Kevin Braga, (401) 578-2036 (cell)

Water Quality

BAL Laboratory, Microbiology (68M00331723)
 Contact: Darlene Capuano, Director, (401) 785-0241

2. R.I. Analytical, Microbiology (68M00331727) Contact: Leslie Koon, (401) 737-8500 ext 109

19. Any samples taken to a lab should include a chain of custody tag, in which all information should be filled out.

- 20. A <u>Fish Kill Investigation Report</u> should be completed and submitted within 24 hours of event, and an electronic copy should be sent to the following staff:
 - Mike Lapisky, Acting Chief, Fish and Wildlife Michael.Lapisky@dem.ri.gov
 - Alicia Good, Assistant Director, Water Resources Alicia.Good@dem.ri.gov
 - Steven Hall, Chief, Enforcement steven.hall@dem.ri.gov
 - Michael Mulhare, Emergency Response <u>mmulhare@dem.state.ri.us</u>
 - Mark Gibson, Deputy Chief, Marine Fisheries mgibson@dem.state.ri.us
 - Najih Lazar, Supervising Biologist, Marine Fisheries najih.lazar@dem.ri.gov

The above procedure has been approved and adopted by

the Office of Marine Fisheries, Division of Fish and Wildlife.

Signature	Title	Date
Signature		Date